



ENVIRONMENT AGENCY

**The Microbiology of Drinking Water (2002) - Part 6 - Methods for the  
isolation and enumeration of sulphite-reducing clostridia  
and *Clostridia perfringens* by membrane filtration**

*Methods for the Examination of Waters and Associated Materials*



## **The Microbiology of Drinking Water (2002) - Part 6 - Methods for the isolation and enumeration of sulphite-reducing clostridia and *Clostridium perfringens* by membrane filtration**

### **Methods for the Examination of Waters and Associated Materials**

This booklet contains two methods for the isolation and enumeration of Sulphite-Reducing Clostridia and *Clostridium perfringens* by membrane filtration

Within this series there are separate booklets dealing with different topics concerning the microbiology of drinking water. Other booklets include

Part 1 - Water quality and public health

Part 2 - Practices and procedures for sampling

Part 3 - Practices and procedures for laboratories

Part 4 - Methods for the isolation and enumeration of coliform bacteria and *Escherichia coli* (including *E. coli* O157:H7)

Part 5 - A method for the isolation and enumeration of enterococci by membrane filtration

Part 7 - Methods for the enumeration of heterotrophic bacteria by pour and spread plate techniques

Part 8 - Methods for the isolation and enumeration of *Aeromonas* and *Pseudomonas aeruginosa* by membrane filtration

Part 9 - Methods for the isolation and enumeration of *Salmonella* and *Shigella* by selective enrichment, membrane filtration and multiple tube most probable number techniques

Part 10 - Methods for the isolation of *Yersinia*, *Vibrio* and *Campylobacter* by selective enrichment

Whilst specific commercial products may be referred to in this document this does not constitute an endorsement of these particular materials. Other similar materials may be suitable and all should be confirmed as such by validation of the method.

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## About this series

### Introduction

This booklet is part of a series intended to provide authoritative guidance on recommended methods of sampling and analysis for determining the quality of drinking water, ground water, river water and sea water, waste water and effluents as well as sewage sludges, sediments and biota. In addition, short reviews of the most important analytical techniques of interest to the water and sewage industries are included.

### Performance of methods

Ideally, all methods should be fully evaluated with results from performance tests. These methods should be capable of establishing, within specified or pre-determined and acceptable limits of deviation and detection, whether or not any sample contains concentrations of parameters above those of interest.

For a method to be considered fully evaluated, individual results from at least three laboratories should be reported. The specifications of performance generally relate to maximum tolerable values for total error (random and systematic errors) systematic error (bias) total standard deviation and limit of detection. Often, full evaluation is not possible and only limited performance data may be available.

In addition, good laboratory practice and analytical quality control are essential if satisfactory results are to be achieved.

### Standing Committee of Analysts

The preparation of booklets within the series "Methods for the Examination of Waters and Associated Materials"

and their continuing revision is the responsibility of the Standing Committee of Analysts. This committee was established in 1972 by the Department of the Environment and is now managed by the Environment Agency. At present, there are nine working groups, each responsible for one section or aspect of water quality analysis. They are

- 1 General principles of sampling and accuracy of results
- 2 Microbiological methods
- 3 Empirical and physical methods
- 4 Metals and metalloids
- 5 General non-metallic substances
- 6 Organic impurities
- 7 Biological methods
- 8 Biodegradability and inhibition methods
- 9 Radiochemical methods

The actual methods and reviews are produced by smaller panels of experts in the appropriate field, in co-operation with the working group and main committee. The names of those members principally associated with these methods are listed at the back of this booklet.

Publication of new or revised methods will be notified to the technical press. An index of methods is available from the Secretary.

Every effort is made to avoid errors appearing in the published text. If, however, any are found, please notify the Secretary.

Dr D Westwood  
*Secretary*

January 2002

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## Warning to users

The analytical procedures described in this booklet should only be carried out under the proper supervision of competent, trained analysts in properly equipped laboratories.

All possible safety precautions should be followed and appropriate regulatory requirements complied with. This should include compliance with the Health and Safety at Work etc Act 1974 and all regulations made under the Act, and the Control of Substances Hazardous to Health Regulations 1999 (SI 1999/437). Where particular or exceptional hazards exist in carrying out the procedures described in this booklet, then specific attention is noted.

Numerous publications are available giving practical details on first aid and laboratory safety. These should be consulted and be readily accessible to all analysts. Amongst such publications are; "Safe Practices in Chemical Laboratories" and "Hazards in the Chemical Laboratory", 1992, produced by the Royal Society of Chemistry; "Guidelines for Microbiological Safety", 1986, Portland Press, Colchester, produced by Member Societies of the Microbiological Consultative Committee; and "Safety Precautions, Notes for Guidance" produced by the Public Health Laboratory Service. Another useful publication is "Good Laboratory Practice" produced by the Department of Health.

## **A Enumeration of sulphite-reducing clostridia by membrane filtration**

### **A1 Introduction**

Tests for sulphite-reducing clostridia play only a subsidiary role in water examination. The organisms form spores which are environmentally resistant and their presence may indicate soil contamination, although some species may grow in deposits, and be associated with corrosion of distribution pipes. *Clostridium perfringens* is a sulphite-reducing species and is associated with faecal contamination. The significance of sulphite-reducing clostridia and *Clostridium perfringens* in water treatment and supply are described elsewhere<sup>(1)</sup> in this series.

### **A2 Scope**

The method is suitable for the examination of drinking waters including samples from all stages of treatment and distribution, and those source waters of moderate turbidity.

Users wishing to employ this method should verify its performance under their own laboratory conditions<sup>(2)</sup>.

### **A3 Definitions**

Sulphite-reducing clostridia are Gram-positive anaerobic spore-forming rods which, in the context of this method, reduce sulphite to sulphide at 37 °C within 24 hours.

### **A4 Principle**

A volume of sample is filtered and the membrane filter placed on the surface of an agar medium containing sulphite and iron(III). The agar medium is then incubated at 37 °C. Sulphite-reducing clostridia usually produce black colonies as a result of the reduction of sulphite to sulphide which reacts with the iron(III) salt. If only a spore count is required then the sample is heat treated at 75 °C prior to filtration in order to kill vegetative bacteria.

### **A5 Limitations**

The method is suitable for most types of aqueous samples except those with high turbidities which tend to block the membrane filter. This will limit the volume of sample that can be filtered. Accumulated deposit on the membrane filter may mask or inhibit the growth of indicator organisms. The maximum number of colonies that should be counted from a single membrane filter is approximately 100.

### **A6 Health and safety**

Media, reagents and bacteria used in this method are covered by the Control of Substances Hazardous to Health Regulations<sup>(3)</sup> and appropriate risk assessments should be made before adopting this method. Standard laboratory microbiology safety procedures should be followed and guidance is given elsewhere<sup>(2)</sup> in this series.

## A7 Apparatus

Standard laboratory equipment should be used which conforms to the performance criteria outlined elsewhere <sup>(2)</sup> in this series. Principally appropriate membrane filtration apparatus and incubators (fan assisted, static temperature) are required. Other items include:

- A7.1 Sterile sample bottles of appropriate volume, made of suitable material, containing sufficient sodium thiosulphate pentahydrate to give a final concentration in the sample of not less than 18 mg/l (for example, 0.1 ml of a 1.8 % m/v solution of Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>.5H<sub>2</sub>O per 100 ml of sample, or equivalent).
- A7.2 Incubator capable of maintaining a temperature of 37 ± 1.0 °C.
- A7.3 Anaerobic jars, or similar equipment, and anaerobic gas-generating system (for generating atmospheres of approximately 90 % hydrogen and 10 % carbon dioxide).
- A7.4 Filtration apparatus, sterile or sterilisable filter funnels, and source of vacuum.
- A7.5 Sterile membrane filters, for example, white, 47 mm diameter cellulose-based, 0.45 µm nominal pore size.
- A7.6 Smooth-tipped forceps.

## A8 Media and reagents

Commercial formulations of these media and reagents may be available, but may possess minor variations in their formulation. The performance of all media and reagents should be verified prior to use in this method. Variations in the preparation and storage of media should also be verified.

### A8.1 *Tryptose sulphite cycloserine agar without egg yolk*<sup>(4, 5)</sup>

Yeast extract	5 g
Tryptose	15 g
Soya peptone	5 g
Sodium metabisulphite	1 g
Iron (III) ammonium citrate	1 g
Agar	14 g
Distilled, deionised or similar grade water	1 litre

Suspend the ingredients in the water and dissolve by heating and stirring. Sterilise the solution by autoclaving at 121 °C for 15 minutes. Allow the medium to cool to 45 - 48 °C. Add 4 ml of a filter-sterilised solution of D-cycloserine in distilled water at a concentration of 100 mg/ml. Mix thoroughly, and dispense into Petri dishes. The final pH of the medium should be 7.6 ± 0.2. Prepared media should be stored in a refrigerator at a temperature between 2 - 8 °C for not more than one week. Petri

dishes, once removed from the refrigerator, should be discarded if not used, and not returned to storage, as the performance of the medium deteriorates.

## A8.2 *Other media*

Standard and commercial formulations of other media and reagents used in this method include Ringer's solution and maximum recovery diluent.

## **A9 Analytical procedure**

### A9.1 *Sample preparation*

The volumes, and dilutions, of samples should be chosen so that the number of colonies to be counted on the membrane filter lies, if possible, between 20 and 80. With some waters, it may be advantageous to filter a selection of different volumes of sample so that the number of colonies on one of the membrane filters is likely to fall within this range. For treated waters, filter 100 ml of the sample. For polluted waters either filter smaller volumes, or dilute the sample with Ringer's solution or maximum recovery diluent before filtration.

If it is the intention to count only the spores of sulphite-reducing clostridia then the sample volume should be heated to 75 °C (for example, in a water bath) and maintained at this temperature for 10 minutes. The temperature should be monitored by placing an appropriate thermometer in a reference bottle containing the same volume of water as the sample being treated.

### A9.2 *Sample processing*

Place the sterile filtration apparatus in position and connect to a source of vacuum, with the stopcock turned off. Remove the funnel and, holding the edge of the membrane filter with sterile smooth-tipped forceps, place a sterile membrane filter, grid-side upwards, onto the porous disc of the filter base. Replace the sterile funnel securely on the filter base. Pour or pipette the required volume of sample into the funnel. When the volume of sample to be filtered is less than 10 ml, add 10 - 20 ml of sterile diluent (for example, quarter-strength Ringer's solution or maximum recovery diluent) to the funnel before addition of the sample. This aids the dispersion of the bacteria over the entire surface of the membrane filter during the filtration. Open the stopcock and apply a vacuum not exceeding 65 kPa (500 mm of mercury) and filter the sample slowly through the membrane filter. Close the stopcock as soon as the sample has been filtered so that as little air as possible is drawn through the membrane filter.

Remove the funnel and transfer the membrane filter carefully to a well-dried tryptose sulphite cycloserine agar Petri dish. Ensure that no air bubbles are trapped between the membrane filter and the medium. 'Rolling' the membrane filter onto the medium minimises the likelihood of air bubbles becoming trapped.

As the spores of sulphite-reducing clostridia are very resilient, funnels that have been used once should be sterilised by autoclaving before being used again. Placing funnels

in a water bath at this stage may not be sufficient to kill spores. If different volumes of the same sample are to be examined, the funnel may be re-used without sterilising the funnel provided that the smallest volume, or highest dilution of the sample, is filtered first. For different samples, take a fresh pre-sterilised funnel and repeat the filtration process. During the filtration of a series of samples, the filter base need not be sterilised unless it becomes contaminated or a membrane filter becomes damaged. When funnels are not in use they should be covered with a sterile lid or a sterile Petri dish lid.

The time between the end of the filtration step and the beginning of the incubation stage should be as short as possible and no longer than 2 hours.

Incubate the Petri dishes at 37 °C in an anaerobic jar or similar system containing an indicator of anaerobiosis and an atmosphere of approximately 90 % hydrogen and 10 % carbon dioxide. Examine the dishes after 24 hours.

### A9.3 *Reading of results*

After incubation, count all black colonies.

### A9.4 *Confirmation tests*

The specificity of tryptose sulphite cycloserine agar is such that confirmation of isolates is not required. Should there be any doubt about the organisms isolated by this method, then colonies may be picked off for Gram staining.

## **A10 Calculations**

### A10.1 Confirmed sulphite-reducing clostridia

The number of confirmed sulphite-reducing clostridia colonies is generally quoted as the number of colonies per 100 ml. Calculate the confirmed count as follows:

$$\text{Confirmed count/100 ml} = \frac{\text{Number of colonies} \times 100}{\text{Volume of sample filtered (ml)}}$$

## **A11 Expression of results**

Counts for sulphite-reducing clostridia are expressed in colony forming units per volume of sample. For drinking water, the volume is typically 100 ml.

## **A12 Quality assurance**

New batches of media and reagents should be tested with appropriate reference strains of target bacteria (for example *Clostridium perfringens*) and non-target bacteria (for example *Bacillus* species). Petri dishes should be incubated for 24 hours at 37 °C. Further details are given elsewhere<sup>(2)</sup> in this series.

## A13 References

1. Standing Committee of Analysts, The Microbiology of Drinking Water (2002) - Part 1 - Water Quality and Public Health. *Methods for the Examination of Waters and Associated Materials*, in this series, Environment Agency.
2. Standing Committee of Analysts, The Microbiology of Drinking Water (2002) - Part 3 - Practices and Procedures for Laboratories. *Methods for the Examination of Waters and Associated Materials*, in this series, Environment Agency.
3. The Control of Substances Hazardous to Health Regulations 1999, Statutory Instrument 1999 No. 437.
4. Enumeration of food-borne *Clostridium perfringens* in egg yolk free tryptose-sulphite-cycloserine agar. *Applied Microbiology*, Hauschild, A. H. W. & Hillsheimer, R., 1974, **27**, 521-526.
5. Membrane filtration enumeration of faecal clostridia and *Clostridium perfringens* in water. *Water Research*, Sartory, D. P., 1986, **20**, 1255-1260.

## **B Enumeration of *Clostridia perfringens* by membrane filtration**

### **B1 Introduction**

Tests for *Clostridium perfringens* play only a subsidiary role in water examination. The organisms form spores which are resistant to environmental stress and can persist in the environment for some time. *Clostridium perfringens* is associated with faecal contamination. If found at a time when other faecal indicator organisms are no longer detectable, the organism may indicate remote or intermittent pollution. The monitoring of *Clostridium perfringens* during water treatment processes may be useful in assessing the performance of such treatment. The significance of *Clostridium perfringens* in water treatment and supply are described elsewhere<sup>(1)</sup> in this series.

### **B2 Scope**

The method is suitable for the examination of drinking waters including samples from all stages of treatment and distribution, and those source waters of moderate turbidity.

Users wishing to employ this method should verify its performance under their own laboratory conditions<sup>(2)</sup>.

### **B3 Definitions**

*Clostridium perfringens* is a Gram-positive anaerobic spore-forming rod which, in the context of this method, reduces sulphite to sulphide at 44 °C within 24 hours.

*Clostridium perfringens* is non-motile, reduces nitrate, ferments lactose and liquefies gelatin.

### **B4 Principle**

A volume of sample is filtered and the membrane filter placed on the surface of an agar medium containing sulphite and iron(III). The agar medium is incubated at 44 °C. *Clostridium perfringens* usually produces black colonies as a result of the reduction of sulphite to sulphide which reacts with the iron(III) salt. If only a spore count is required, then the sample is heat treated at 75 °C prior to filtration in order to kill vegetative bacteria.

### **B5 Limitations**

The method is suitable for most types of aqueous samples except those with high turbidities which tend to block the membrane filter. This will limit the volume of sample that can be filtered. Accumulated deposit on the membrane filter may mask or inhibit the growth of indicator organisms. The maximum number of colonies that should be counted from a single membrane is approximately 100.

### **B6 Health and safety**

Media, reagents and bacteria used in this method are covered by the Control of Substances Hazardous to Health Regulations<sup>(3)</sup> and appropriate risk assessments

should be made before adopting this method. Standard laboratory microbiology safety procedures should be followed and guidance is given elsewhere<sup>(2)</sup> in this series.

## **B7 Apparatus**

Standard laboratory equipment should be used which conforms to the performance criteria outlined elsewhere<sup>(2)</sup> in this series. Principally appropriate membrane filtration apparatus and incubators (fan assisted, static temperature) are required. Other items include:

- B7.1 Sterile sample bottles of appropriate volume, made of suitable material, containing sufficient sodium thiosulphate pentahydrate to give a final concentration in the sample of not less than 18 mg/l (for example, 0.1 ml of a 1.8 % m/v solution of Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>·5H<sub>2</sub>O per 100 ml of sample, or equivalent).
- B7.2 Incubators capable of maintaining temperatures of 37 ± 1.0 °C and 44 ± 0.5 °C.
- B7.3 Anaerobic jars, or similar equipment, and anaerobic gas-generating system (for generating atmospheres of approximately 90 % hydrogen and 10 % carbon dioxide).
- B7.4 Filtration apparatus, sterile or sterilisable filter funnels, and source of vacuum.
- B7.5 Sterile membrane filters, for example, white, 47 mm diameter, cellulose-based, 0.45 µm nominal pore size.
- B7.6 Smooth-tipped forceps.

## **B8 Media and reagents**

Commercial formulations of these media and reagents may be available, but may possess minor variations in their formulation. The performance of all media and reagents should be verified prior to use in this method. Variations in preparation and storage of media should also be verified.

### **B8.1 *Tryptose sulphite cycloserine agar without egg yolk* <sup>(4, 5)</sup>**

Yeast extract	5 g
Tryptose	15 g
Soya peptone	5 g
Sodium metabisulphite	1 g
Iron (III) ammonium citrate	1 g
Agar	14 g
Distilled, deionised or similar grade water	1 litre

Suspend the ingredients in the water and dissolve by heating and stirring. Sterilise the solution by autoclaving at 121 °C for 15 minutes. Allow the medium to cool to 45 - 48 °C. Add 4 ml of a filter-sterilised solution of D-cycloserine in distilled water at a concentration of 100 mg/ml. Mix thoroughly, and dispense into Petri dishes. The

final pH of the medium should be  $7.6 \pm 0.2$ . Prepared medium should be stored in a refrigerator at a temperature between 2 - 8 °C, for not more than one week. Petri dishes, once removed from the refrigerator should be discarded if not used, and not returned to storage, as the performance of the medium deteriorates.

#### B8.2 *Buffered nitrate-motility medium*<sup>(6)</sup>

Beef extract	3 g
Peptone	5 g
Potassium nitrate	5 g
D-Galactose	5 g
Glycerol	5 g
Disodium hydrogen phosphate	2.5 g
Agar	3 g
Distilled, deionised or similar grade water	1 litre

Dissolve the ingredients in the water and dispense in 10 ml aliquots in appropriately sized capped tubes. Sterilise the medium by autoclaving at 121 °C for 15 minutes. The final pH of the medium should be  $7.3 \pm 0.2$ . Prepared tubes should be stored at a temperature between 2 - 8 °C for up to one month if protected against dehydration.

#### B8.3 *Nitrate reduction test reagents*

Reagent A	
Sulphanilic acid	0.8 g
5N acetic acid	100 ml

Warm gently to aid dissolving.

Reagent B	
N, N-dimethyl $\alpha$ -naphthylamine	0.6 g
5N acetic acid.	100 ml

Dissolve the amine in the acetic acid solution.

The reagents should be stored at a temperature between 2 - 8 °C.

#### B8.4 *Lactose-gelatin medium*<sup>(4)</sup>

Tryptose	15 g
Yeast extract	10 g
Disodium hydrogen phosphate	5 g
Gelatin	120 g
Lactose	10 g
Phenol red (0.4 % m/v solution)	12.5 ml
Distilled, deionised or similar grade water	1 litre

Dissolve the ingredients, except the lactose and phenol red, in the water and adjust the pH to  $7.5 \pm 0.2$ . Add the lactose and phenol red and dissolve. Dispense in 10 ml

aliquots in appropriately sized capped tubes and sterilise the medium at 121 °C for 15 minutes. The final pH should be  $7.5 \pm 0.2$ . Prepared media may be stored at a temperature between 2 - 8 °C for up to one month, if protected against dehydration.

#### B8.5 *Other media*

Standard and commercial formulations of other media and reagents used in this method include Ringer's solution and maximum recovery diluent.

### **B9 Analytical procedure**

#### B9.1 *Sample preparation*

The volumes, and dilutions, of samples should be chosen so that the number of colonies to be counted on the membrane filter lies, if possible, between 20 and 80. With some waters, it may be advantageous to filter a selection of different volumes of sample so that the number of colonies on one of the membrane filters is likely to fall within this range. For treated waters, filter 100 ml of the sample. For polluted waters either filter smaller volumes, or dilute the sample with Ringer's solution or maximum recovery diluent before filtration.

If it is the intention to count only the spores of *Clostridium perfringens* then the volume of sample should be heated to 75 °C (for example in a water bath) and maintained at this temperature for 10 minutes. The temperature should be monitored by placing an appropriate thermometer in a reference bottle containing the same volume of water as the sample being treated.

#### B9.2 *Sample processing*

Place the sterile filtration apparatus in position and connect to a source of vacuum, with the stopcock turned off. Remove the funnel and, holding the edge of the membrane filter with sterile smooth-tipped forceps, place a sterile membrane filter, grid-side upwards, on the porous disc of the filter base. Replace the sterile funnel securely on the filter base. Pour or pipette the required volume of sample into the funnel. When the volume of sample to be filtered is less than 10 ml, add 10 - 20 ml of sterile diluent (for example, quarter-strength Ringer's solution or maximum recovery diluent) to the funnel before addition of the sample. This aids dispersion of the bacteria over the entire surface of the membrane filter during filtration. Open the stopcock and apply a vacuum not exceeding 65 kPa (500 mm of mercury) and filter the sample slowly through the membrane filter. Close the stopcock as soon as the sample has been filtered so that as little air as possible is drawn through the membrane filter.

Remove the funnel and transfer the membrane filter carefully to a well-dried tryptose sulphite cycloserine agar Petri dish. Ensure that no air bubbles are trapped between the membrane filter and the medium. 'Rolling' the membrane filter onto the medium minimises the likelihood of air bubbles becoming trapped.

As the spores of *Clostridium perfringens* are very resilient, funnels that have been used once should be sterilised by autoclaving before being used again. Placing funnels in a water bath at this stage may not be sufficient to kill spores. If different volumes of the same sample are to be examined, the funnel may be re-used without sterilising the funnel provided that the smallest volume, or highest dilution of sample, is filtered first. For different samples, take a fresh pre-sterilised funnel and repeat the filtration process. During the filtration of a series of samples, the filter base need not be sterilised unless it becomes contaminated or a membrane filter becomes damaged. When funnels are not in use they should be covered with a sterile lid or a sterile Petri dish lid.

The time between the end of the filtration step and the beginning of the incubation stage should be as short as possible and no longer than 2 hours.

Incubate the Petri dishes at 44 °C in an anaerobic jar or similar system containing an indicator of anaerobiosis and an atmosphere of approximately 90 % hydrogen and 10 % carbon dioxide. Examine the dishes after 24 hours. It has been reported that to ensure anaerobic conditions are satisfied, the use of an agar overlay results in higher numbers of colonies than when the membrane filter is incubated on the surface of the agar.

### B9.3 *Reading of results*

After incubation, count all black colonies.

### B9.4 *Confirmation tests*

Depending on the intended purpose of the analysis and the required accuracy, sub-culture a suitable number of black colonies (however faint). If the aim is to estimate the number of organisms present, then for the greatest accuracy, all colonies should be sub-cultured if fewer than ten are present or, at least ten colonies should be sub-cultured if more than ten are present.

For each isolate, inoculate a tube of buffered nitrate-motility medium and incubate anaerobically at 37 °C for 24 hours. To test for nitrate reduction, add 1 ml of nitrate test reagent A and 1 ml of nitrate test reagent B to each tube. A red colour indicates nitrate reduction to nitrite.

In addition, inoculate a tube of lactose-gelatin medium and incubate anaerobically at 37 °C for 48 hours. After incubation, the tubes are placed in a refrigerator for at least one hour before observing whether any gelatin liquefaction has occurred.

*Clostridium perfringens* is confirmed by the following reactions:

- (i) Non-motile - growth not spread through buffered nitrate-motility medium.
- (ii) Nitrate reducing - red colour on addition of nitrate test reagents A and B to buffered nitrate-motility medium.

- (iii) Lactose fermenting - yellow colouration of lactose-gelatin medium.
- (iv) Gelatin liquefying - contents of the lactose-gelatin medium tube become liquefied.

## **B10 Calculations**

### **B10.1 Presumptive *Clostridium perfringens***

The number of presumptive *Clostridium perfringens* colonies is generally expressed as the number of colonies per 100 ml. Calculate the presumptive count as follows:

$$\text{Presumptive count/100 ml} = \frac{\text{Number of colonies} \times 100}{\text{Volume of sample filtered (ml)}}$$

### **B10.2 Confirmed *Clostridium perfringens***

The number of confirmed *Clostridium perfringens* colonies is calculated by multiplying the number of presumptive *Clostridium perfringens* by the proportion of the isolates that are non-motile, reduce nitrate, ferment lactose and liquefy gelatin.

## **B11 Expression of results**

The number of presumptive and confirmed *Clostridium perfringens* is expressed in colony forming units per volume of sample. For drinking waters, the volume is typically 100 ml.

## **B12 Quality assurance**

New batches of media and reagents should be tested with appropriate reference strains of target bacteria (for example *Clostridium perfringens*) and non-target bacteria (for example *Bacillus* species). Petri dishes should be incubated for 24 hours at 37 °C or 44 °C as appropriate. Further details are given elsewhere<sup>(2)</sup> in this series.

## **B13 References**

1. Standing Committee of Analysts, The Microbiology of Drinking Water (2002) - Part 1 - Water Quality and Public Health. *Methods for the Examination of Waters and Associated Materials*, in this series, Environment Agency.
2. Standing Committee of Analysts, The Microbiology of Drinking Water (2002) - Part 3 - Practices and Procedures for Laboratories. *Methods for the Examination of Waters and Associated Materials*, in this series, Environment Agency.
3. The Control of Substances Hazardous to Health Regulations 1999, Statutory Instrument 1999 No. 437.

4. Enumeration of food-borne *Clostridium perfringens* in egg yolk free tryptose-sulphite-cycloserine agar. *Applied Microbiology*, Hauschild, A. H. W. & Hillsheimer, R., 1974, **27**, 521-526.
5. Membrane filtration enumeration of faecal clostridia and *Clostridium perfringens* in water. *Water Research*, Sartory, D. P., 1986, **20**, 1255-1260.
6. Media for confirming *Clostridium perfringens* from food and faeces. *Journal of Food Protection*, Harmon, S. M. & Kautter, D. A., 1978, **41**, 626-630.

## **Address for correspondence**

However well procedures may be tested, there is always the possibility of discovering hitherto unknown problems. Analysts with such information are requested to contact the Secretary of the Standing Committee of Analysts at the address given below.

Secretary  
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## **Environment Agency Standing Committee of Analysts Members assisting with these methods**

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Solihull B91 1QT  
Tel: 0121 711 2324  
Fax: 0121 711 5824

#### SOUTH WEST

Manley House  
Kestrel Way  
Exeter EX2 7LQ  
Tel: 01392 444 000  
Fax: 01392 444 238

#### NORTH EAST

Rivers House  
21 Park Square South  
Leeds LS1 2QG  
Tel: 0113 244 0191  
Fax: 0113 246 1889

#### THAMES

Kings Meadow House  
Kings Meadow Road  
Reading RG1 8DQ  
Tel: 0118 953 5000  
Fax: 0118 950 0388

#### NORTH WEST

PO Box 12  
Richard Fairclough House  
Knutsford Road  
Warrington WA4 1HG  
Tel: 01925 653 999  
Fax: 01925 415 961

#### WALES

Rivers House/Plas-yr-Afon  
St Mellons Business Park  
Fortran Road  
St Mellons  
Cardiff CF3 0EY  
Tel: 029 2077 0088  
Fax: 029 2079 8555



ENVIRONMENT AGENCY  
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**0845 988 1188**

ENVIRONMENT AGENCY  
EMERGENCY HOTLINE

**0800 80 70 60**



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